

The Hyperglycemia Stranglehold Stifles Cutaneous Epithelial–Mesenchymal Plasticity and Functional Wound Closure



Chandan K. Sen¹ and Sashwati Roy¹

Iterative cycles of epithelial–mesenchymal transition (EMT) and mesenchymal-to-epithelial transition (MET) are responsible for epithelial plasticity necessary to achieve functional wound closure. Restoration of the barrier function of the repaired skin is a hallmark of functional wound closure. Both EMT and MET are subject to control by glycemic status. A new article by Tan et al (2020) supports the notion that hyperglycemia blunts epithelial plasticity.

Journal of Investigative Dermatology (2021) **141**, 1382–1385. doi:10.1016/j.jid.2020.11.021

Skin wound repair, regenerative and near perfect under fetal conditions, remains intact with scarring in healthy adults but stalls under diabetic conditions (Gnyawali et al., 2020). During morphogenesis, epithelial cells dismantle cell adhesion and tight junction structures in an effort to acquire a mesenchymal phenotype. This process, termed epithelial–mesenchymal transition (EMT), is prominent in fetal skin. It is noted after epidermal development, and it gives rise to dermal α -smooth muscle actin–expressing cells. In adults, these cells contribute to wound contraction and re-epithelialization resulting in wound closure with a characteristic scar phenotype (Kong et al., 2006). EMT is marked by the induction of prototypic epithelial markers coupled with the loss of apical–basal polarity and increased cell motility caused by cytoskeleton reorganization. Re-epithelialization of wounds relies on turning down intercellular adhesion followed by keratinocyte (KC) migration in the epidermis proximal to wound margins. Purse-string wound contraction caused by EMT-derived myofibroblasts at the same time prepares the underlying

connective tissue bed. Now, Tan et al. (2020) report that hyperglycemia restrains KC EMT. Specifically, their work shows that acetylcholine (ACh)-induced EMT is at risk under conditions of hyperglycemia jeopardizing diabetic wound repair (Tan et al., 2020). This work draws attention to the significance of non-neuronal ACh in the regulation of diabetic wound healing. Cholinergic pathways and nicotinic and muscarinic receptors are known to be present in KCs. Impairment in these receptors can cause Grover disease, an eruption of intraepidermal acantholysis presenting as crusted reddened papules (Paslin, 2012). In vitro studies show that ACh improves cell migration (Uberti et al., 2017), and in vivo, cholinergic peptides augment skin wound closure (Chernyavsky et al., 2012).

Covering of wounds without discharge is an inadequate marker of wound closure because biofilm-infected wounds may achieve such closure without restoring the barrier function of the repaired skin. Repair that results in barrier function–deficient skin is faulty because it compromises the biomechanical properties of closed wounds in a way that favors wound

recurrence. The concept of functional wound closure has thus emerged. Functional wound closure is achieved when covering of the wound defect is achieved without discharge and with evidence of restored barrier function at the site of closure (Roy et al., 2020, 2014). Of interest in this context, cholinergic pathways in KCs induce the production of antimicrobial peptides and improve the barrier function of the skin (Curtis and Radek, 2012). In wound healing, EMT influences vascularization as well as re-epithelialization. Cutaneous EMT regulates wound angiogenesis and closure in a glycemic status–dependent manner (Singh et al., 2019). Stalled wound re-epithelialization and compromised angiogenesis are hallmarks of impaired diabetic wound healing. Work by our group recently identified ZEB1 as a significant mechanistic hub across epithelial and endothelial cells in wounds. In both epithelial as well as endothelial cell compartments of wound tissue, ZEB1 is responsive to the glycemic status of the injury microenvironment. In epithelial cells, hyperglycemia impaired the ZEB1-EMT pathway toward wound epithelialization. In endothelial cells, ZEB1 was directly implicated in hyperglycemia-induced dysfunction (Singh et al., 2019). These findings establish a direct link of EMT with critical facets of wound healing, including functional closure and wound-site vascularization.

Wound closure is only complete when defects caused by injury are covered by skin with a restored barrier function. Thus, measurement of barrier function restoration is an important element characterizing wound closure (Ghatak et al., 2015; Li et al., 2018a). Inherent reversible plasticity of skin cells is manifest during wound repair (Figure 1). One aspect of this process is EMT ↔ mesenchymal-to-epithelial transition (MET) (Lamouille et al., 2014; Nieto et al., 2016). The spatiotemporal process that advances re-epithelialization in a way that restores epidermal barrier function requires partial EMT (Haensel and Dai, 2018). Extracellular cues trigger transcriptional, translational, and post-translational regulation of transcription

¹Indiana Center for Regenerative Medicine and Engineering, Indiana University Health Comprehensive Wound Center, Department of Surgery, Indiana University School of Medicine, Indianapolis, Indiana, USA

Correspondence: Chandan K. Sen, Indiana Center for Regenerative Medicine and Engineering, Suite 454 IB, 975 West Walnut Street, Indianapolis, Indiana 46202, USA. E-mail: cksen@iu.edu

© 2021 The Authors. Published by Elsevier, Inc. on behalf of the Society for Investigative Dermatology.

Clinical Implications

- Management of hyperglycemia will defend skin plasticity necessary for wound closure.
- Management of hyperglycemia will enable cholinergic pathways of the skin to support wound closure.
- Management of hyperglycemia will help achieve functional wound closure.

factors (TFs) such as SNAIL, ZEB, and basic helix-loop-helix TF to cause EMT. TGF β family signaling pathways play a central role in relaying those cues. Of the many signaling pathways that enable EMT, the significance of non-neuronal cholinergic pathways in driving EMT in the skin remains poorly understood (Lamouille et al., 2014). Tan et al. (2020)

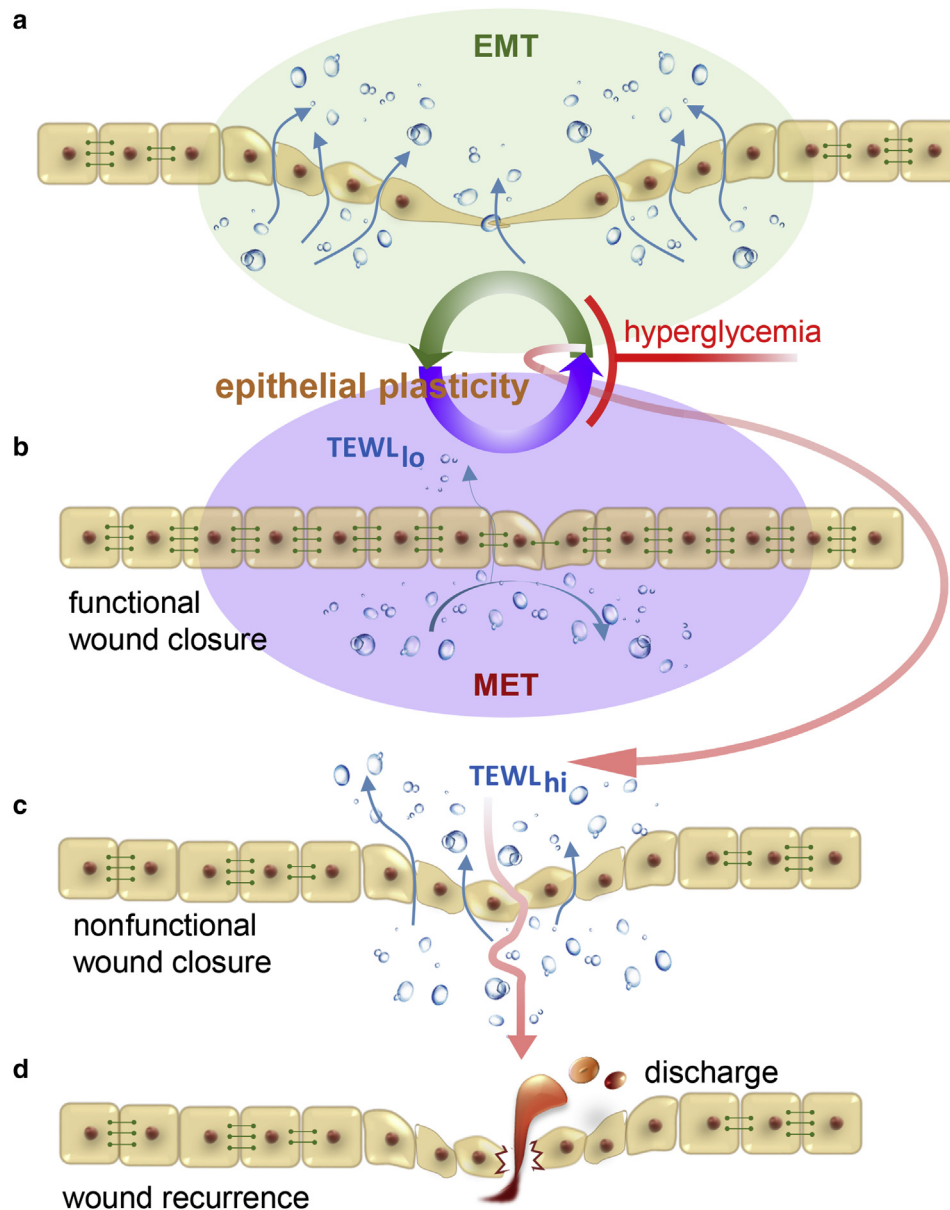


Figure 1. Hyperglycemia restrains cutaneous epithelial plasticity necessary for functional wound closure. A hypothetical paradigm depicts (a) and (b) in which EMT \leftrightarrow MET is central for re-epithelialization and restoration of the barrier function of the repaired skin. During EMT, epithelial cells dismantle cell adhesion and tight junction structures in an effort to acquire a mesenchymal phenotype favoring re-epithelialization. MET helps to reconstitute the AJCs restoring the barrier function of the repaired skin. (c) Hyperglycemia is a barrier to such plasticity and as such hinders re-epithelialization, restoration of barrier function, or both. The result is wound chronicity or nonfunctional wound closure. (d) Closure of wound without the restoration of skin barrier function predisposes the closed wound to recidivism as evident in preclinical porcine studies. The incidence of wound recurrence is high in patients with diabetes. TEWL is a measure of skin barrier function. TEWL_{hi} represents TEWL-deficient skin barrier function. TEWL_{lo} represents TEWL-restored skin barrier function indicative of functional wound closure. Horizontal rivets between cells represent functional AJC. These are low or absent in mesenchymal cells compromising barrier function. AJC, apical junctional complex; EMT, epithelial-to-mesenchymal transition; MET, mesenchymal-to-epithelial transition; TEWL, transepidermal water loss.

COMMENTARY

report that under conditions of hyperglycemia, KCs were resistant to ACh-induced EMT. ACh-dependent EMT was dependent on TGF β 1 signaling. In the skin, the activation of the parasympathetic nervous system by stimuli, including stress, causes the release of ACh from nerve fibers. Among other outcomes, this causes sweating. In those suffering from wounds, stress is commonly experienced (Sen and Roy, 2019). Thus, it is plausible that ACh is present in elevated levels at sites of cutaneous wounds. Both ACh-synthesizing choline acetyltransferase as well as the ACh-degrading enzyme acetylcholinesterase are abundant in the skin. On the basis of the work by Tan et al. (2020), it may be that ACh represents a physiological mechanism to augment EMT in wounds.

Central cholinergic pathways have a profound influence on glycemic regulation (Healy et al., 2010). However, information on how hyperglycemia may modify cholinergic responses in peripheral organs is scanty. Type 2 diabetes blunts purinergic cutaneous vasodilatation but not muscarinic and nicotinic vascular responses or sweating (Fujii et al., 2018). Kevin Tracey's (Tracey, 2009) cholinergic anti-inflammatory pathway provides important context to the work reported by Tan et al. (2020). In an effort to understand the anti-inflammatory effects of a p38 MAPK inhibitor, these investigators identified an inflammatory reflex. In brief, it was proposed that the vagus nerve can sense peripheral inflammation and, in response, dispatch action potentials aimed at inhibiting proinflammatory cytokine production by the spleen. As part of molecular mechanisms that drive the cholinergic anti-inflammatory pathway, neurotransmitter ACh acts on the α 7 nicotinic ACh receptor (α 7nAChR) subunit expressed on cytokine-producing cells such as monocytes, macrophages, and lymphocytes (Huston and Tracey, 2011). Neuroendocrine α 7nAChR is also functionally active in skin cells such as epidermal KCs, sebocytes, and dermal fibroblasts. Both successful mounting and timely resolution of inflammation are necessary for wound healing (Khanna et al., 2010). Diabetic wound repair is complicated by persistent inflammation. Among several other factors (Das et al., 2018, 2016, 2015,

2014), α 7nAChR function is likely to play a considerable role in this regard. Selective agonists of α 7nAChR accelerated the repair of diabetic wounds (Li et al., 2018b). nAChRs also accelerate diabetic wound angiogenesis (Jacobi et al., 2002). In diabetes, α 7nAChR expression and function are blunted. Receptor for advanced glycation end products inactivates α 7nAChR (Chandna et al., 2015). In the context of wound closure, it is important to note that α 7nAChR is directly implicated in driving EMT (Zhang et al., 2016; Zhao et al., 2015). Whether diabetes-dependent impairment of α 7nAChR function in skin cells impairs EMT during wound closure remains an open question.

Once re-epithelialization is achieved, epithelial cells must give up their migratory behavior, reconstitute apico-basal polarization, and re-establish junctional complexes. Cells must reverse EMT, and this can be achieved by MET (Thiery et al., 2009). Restoring the barrier function of repaired skin requires the reconstitution of apical junctional complexes encompassing tight junctions and adherens junctions. Although the specifics of EMT \leftrightarrow MET mechanisms during wound healing have not been worked out yet, the mechanisms may reasonably be hypothesized to be an iterative process. Importantly, the same inducer can potentiate EMT and MET simultaneously in two different cell compartments. If clues from the formation of complex three-dimensional structures of internal organs are of any value, several rounds of EMT and MET are necessary for the final differentiation of specialized cell types (Thiery et al., 2009). Iterative EMT \leftrightarrow MET may be viewed as stepwise cycles of epithelial plasticity necessary to achieve functional wound closure. Of extraordinary significance in the context of diabetic wounds is the evidence that tissue EMT \leftrightarrow MET is responsive to glycemic status (Talakatta et al., 2018). Further studies unveiling the molecular underpinnings of cutaneous wound epithelial plasticity will reveal regulatory hubs orchestrating wound inflammation, re-epithelialization, and vascularization.

ORCIDiS

Chandan K. Sen: <http://orcid.org/0000-0003-3151-5202>

Sashwati Roy: <http://orcid.org/0000-0002-9995-4917>

CONFLICT OF INTEREST

The authors state no conflict of interest.

ACKNOWLEDGMENT

Wound healing research in the authors' laboratories is supported by DK119099, DK125835, NR015676, NS042617, and DK114718.

REFERENCES

- Chandna AR, Nair M, Chang C, Pennington PR, Yamamoto Y, Mousseau DD, et al. RAGE mediates the inactivation of nAChRs in sympathetic neurons under high glucose conditions. *Eur J Neurosci* 2015;41:341–51.
- Chernyavsky AI, Kalantari-Dehaghi M, Phillips C, Marchenko S, Grando SA. Novel cholinergic peptides SLURP-1 and -2 regulate epithelialization of cutaneous and oral wounds. *Wound Repair Regen* 2012;20:103–13.
- Curtis BJ, Radek KA. Cholinergic regulation of keratinocyte innate immunity and permeability barrier integrity: new perspectives in epidermal immunity and disease. *J Invest Dermatol* 2012;132:28–42.
- Das A, Datta S, Roche E, Chaffee S, Jose E, Shi L, et al. Novel mechanisms of Collagenase Santyl Ointment (CSO) in wound macrophage polarization and resolution of wound inflammation. *Sci Rep* 2018;8:1696.
- Das A, Ganesh K, Khanna S, Sen CK, Roy S. Engulfment of apoptotic cells by macrophages: a role of microRNA-21 in the resolution of wound inflammation. *J Immunol* 2014;192:1120–9.
- Das A, Ghatak S, Sinha M, Chaffee S, Ahmed NS, Parinandi NL, et al. Correction of MFG-E8 resolves inflammation and promotes cutaneous wound healing in diabetes. *J Immunol* 2016;196:5089–100.
- Das A, Sinha M, Datta S, Abas M, Chaffee S, Sen CK, et al. Monocyte and macrophage plasticity in tissue repair and regeneration. *Am J Pathol* 2015;185:2596–606.
- Fujii N, Meade RD, McNeely BD, Nishiyasu T, Sigal RJ, Kenny GP. Type 2 diabetes specifically attenuates purinergic skin vasodilatation without affecting muscarinic and nicotinic skin vasodilatation and sweating. *Exp Physiol* 2018;103:212–21.
- Ghatak S, Chan YC, Khanna S, Banerjee J, Weist J, Roy S, et al. Barrier function of the repaired skin is disrupted following arrest of dicer in keratinocytes. *Mol Ther* 2015;23:1201–10.
- Gnyawali SC, Sinha M, El Masry MS, Wulff B, Ghatak S, Soto-Gonzalez F, et al. High resolution ultrasound imaging for repeated measure of wound tissue morphometry, biomechanics and hemodynamics under fetal, adult and diabetic conditions. *PLoS One* 2020;15:e0241831.
- Haensel D, Dai X. Epithelial-to-mesenchymal transition in cutaneous wound healing: where we are and where we are heading. *Dev Dyn* 2018;247:473–80.
- Healy JA, Nilsson KR, Hohmeier HE, Berglund J, Davis J, Hoffman J, et al. Cholinergic augmentation of insulin release requires ankyrin-B. *Sci Signal* 2010;3:ra19.
- Huston JM, Tracey KJ. The pulse of inflammation: heart rate variability, the cholinergic anti-

- inflammatory pathway and implications for therapy. *J Intern Med* 2011;269:45–53.
- Jacobi J, Jang JJ, Sundram U, Dayoub H, Fajardo LF, Cooke JP. Nicotine accelerates angiogenesis and wound healing in genetically diabetic mice. *Am J Pathol* 2002;161:97–104.
- Khanna S, Biswas S, Shang Y, Collard E, Azad A, Kauh C, et al. Macrophage dysfunction impairs resolution of inflammation in the wounds of diabetic mice. *PLoS One* 2010;5:e9539.
- Kong W, Li S, Liu C, Bari AS, Longaker MT, Lorenz HP. Epithelial-mesenchymal transition occurs after epidermal development in mouse skin. *Exp Cell Res* 2006;312:3959–68.
- Lamouille S, Xu J, Derynck R. Molecular mechanisms of epithelial-mesenchymal transition. *Nat Rev Mol Cell Biol* 2014;15:178–96.
- Li J, Ghatak S, El Masry MS, Das A, Liu Y, Roy S, et al. Topical lyophilized targeted lipid nanoparticles in the restoration of skin barrier function following burn wound. *Mol Ther* 2018;26:2178–88.
- Li JY, Jiang SK, Wang LL, Zhang MZ, Wang S, Jiang ZF, et al. $\alpha 7$ -nAChR activation has an opposite effect on healing of covered and uncovered wounds. *Inflammation* 2018;41:474–84.
- Nieto MA, Huang RY, Jackson RA, Thiery JP. EMT. 2016. *Cell* 2016;166:21–45.
- Paslin D. Grover disease may result from the impairment of keratinocytic cholinergic receptors. *J Am Acad Dermatol* 2012;66:332–3.
- Roy S, Elgharably H, Sinha M, Ganesh K, Chaney S, Mann E, et al. Mixed-species biofilm compromises wound healing by disrupting epidermal barrier function. *J Pathol* 2014;233:331–43.
- Roy S, Santra S, Das A, Dixith S, Sinha M, Ghatak S, et al. Staphylococcus aureus biofilm infection compromises wound healing by causing deficiencies in granulation tissue collagen. *Ann Surg* 2020;271:1174–85.
- Sen CK, Roy S. Sociogenomic approach to wound care: a new patient-centered paradigm. *Adv Wound Care (New Rochelle)* 2019;8:523–6.
- Singh K, Sinha M, Pal D, Tabasum S, Gnyawali SC, Khona D, et al. Cutaneous epithelial to mesenchymal transition activator ZEB1 regulates wound angiogenesis and closure in a glycemic status-dependent manner. *Diabetes* 2019;68:2175–90.
- Talakatta G, Sarikhani M, Muhamed J, Dhanya K, Somashekar BS, Mahesh PA, et al. Diabetes induces fibrotic changes in the lung through the activation of TGF- β signaling pathways. *Sci Rep* 2018;8:11920.
- Tan MWY, Tan WR, Kong ZQ, Toh JH, Wee WKJ, Teo EML, et al. High Glucose Restraint of Acetylcholine-Induced Keratinocyte Epithelial-Mesenchymal Transition Is Mitigated by p38 Inhibition. *J Invest Dermatol* 2021;141:1438–49.e9.
- Thiery JP, Acloque H, Huang RY, Nieto MA. Epithelial-mesenchymal transitions in development and disease. *Cell* 2009;139:871–90.
- Tracey KJ. Reflex control of immunity. *Nat Rev Immunol* 2009;9:418–28.

Uberti F, Bardelli C, Morsanuto V, Ghirlanda S, Cochis A, Molinari C. Stimulation of the nonneuronal cholinergic system by highly diluted acetylcholine in keratinocytes. *Cells Tissues Organs* 2017;203:215–30.

Zhang C, Ding XP, Zhao QN, Yang XJ, An SM, Wang H, et al. Role of $\alpha 7$ -nicotinic acetylcholine receptor in nicotine-induced invasion and

epithelial-to-mesenchymal transition in human non-small cell lung cancer cells. *Oncotarget* 2016;7:59199–208.

Zhao Q, Gu X, Zhang C, Lu Q, Chen H, Xu L. Blocking M2 muscarinic receptor signaling inhibits tumor growth and reverses epithelial-mesenchymal transition (EMT) in non-small cell lung cancer (NSCLC). *Cancer Biol Ther* 2015;16:634–43.

See related article on pg 1553

The Enigma of AHR Activation in the Skin: Interplay among Ligands, Metabolism, and Bioavailability

Ellen H. van den Bogaard¹ and Gary H. Perdeu²

AHR is expressed in a variety of skin cell types and contributes to skin homeostasis. Therapeutic targeting of AHR in dermatology was first described for the treatment of inflammatory skin diseases using coal tar ointment. In addition to therapies involving active ligands such as coal tar and tapinarof, the inhibition of AHR-dependent enzymatic activities in the skin may be an alternative approach to resolving skin inflammation.

Journal of Investigative Dermatology (2021) **141**, 1385–1388. doi:10.1016/j.jid.2020.12.013

Introduction

Ten years ago, AHR, a ligand-activated transcription factor, was best known for its role in mediating toxicity and carcinogenesis through the binding of environmental pollutants and xenobiotics. Recently, this promiscuous environmental sensor has gained tremendous attention as a modulator of immune cell development and immune tolerance in a tissue-dependent manner. In the fields of skin biology and dermatology, research into AHR signaling is booming. Multiple studies describe the efficacy of exogenous ligands to induce terminal differentiation, increase the expression levels of skin barrier-related and antimicrobial proteins, and thus contribute to skin barrier function. AHR activation also

interferes with cytokine-induced or T-cell-mediated inflammatory signaling pathways (e.g., through Jak/signal transducer and activator of transcription) (Furie, 2020). These anti-inflammatory effects lead to reduced symptoms in experimental models of psoriasis and atopic dermatitis (AD), similar to the well-known therapeutic efficacy and molecular mechanism of action of coal tar in the treatment of these chronic inflammatory skin diseases (van den Bogaard et al., 2013). It is important to keep in mind that improvement of epithelial barrier function would result in a dampening of inflammation, and, thus, these two proposed functions of the AHR in the skin would be linked. The accumulating body of evidence on the role of AHR

¹Laboratory for Experimental Dermatology, Department of Dermatology, Radboud Institute for Molecular Life Sciences, Radboud University Medical Center, Nijmegen, The Netherlands; and ²Center for Molecular Toxicology and Carcinogenesis, Department of Veterinary and Biomedical Sciences, The Pennsylvania State University, University Park, Pennsylvania, USA

Correspondence: Ellen H. van den Bogaard, Department of Dermatology, Radboud Institute for Molecular Life Sciences, Radboud University Medical Center, Post Office Box 9101, 6500 HB Nijmegen, The Netherlands. E-mail: ellen.vandenbogaard@radboudumc.nl

© 2021 The Authors. Published by Elsevier, Inc. on behalf of the Society for Investigative Dermatology.

