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Rapid Letter

Physical Exercise Induces Activation of NF- κ B in Human Peripheral Blood Lymphocytes

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ABSTRACT

Current understanding of nuclear factor- κ B (NF- κ B) activation is derived mostly from *in vitro* studies, and *in vivo* human data are limited. This study provides first evidence showing that physical exercise (80% maximal O₂ consumption, 1 h) may trigger NF- κ B activation, as determined by electrophoretic mobility shift assay, in peripheral blood lymphocytes of physically fit young men. Supershift assay showed that the NF- κ B protein complex contained the transcriptionally active p65 protein. Plasma levels of NF- κ B-directed gene products such as tumor necrosis factor- α and interleukin-2 receptor confirmed that physical exercise caused NF- κ B transactivation. Exercise-induced NF- κ B activation in lymphocytes was associated with elevated levels of lipid peroxidation by-products in the plasma. *Antioxid. Redox Signal.* 3, 1131–1137.

INTRODUCTION

THE NUCLEAR FACTOR- κ B (NF- κ B) p50/p65 heterodimer is the classical member of the Rel family of transcription factors that regulate diverse cellular functions such as immune response, cell growth, survival, and development (2, 3, 7, 33). In lymphocytes, NF- κ B activation represents a normal component of cell response to a wide variety of stimuli (2, 6, 16, 28, 30). NF- κ B regulates cell division, apoptosis, and differentiation that accompany lymphocyte activation (6). NF- κ B activity is induced by a wide variety of factors, including cytokines, phosphatase inhibitors, endotoxin and certain protein phosphorylation agonists (2, 3). Oxidants have been

shown to induce NF- κ B activation in several types, and it has been postulated that oxidants may serve as intracellular messengers inducing NF- κ B activity (14, 21, 30, 33).

Strenuous or prolonged exercise is known to induce oxidative stress (29, 31, 32). In humans, physical exercise has been observed to induce oxidative DNA damage in lymphocytes (11). It has been suggested that neutrophil-derived oxidants cause such damage (22). Most of our current knowledge regarding the various factors that induce NF- κ B activity is derived from *in vitro* studies (2, 3, 7). Information from human *in vivo* studies is limited. In this study, we sought to investigate whether strenuous prolonged exercise influences lymphocyte NF- κ B activity in physically fit men.

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MATERIALS AND METHODS

Subjects

Twelve healthy male endurance and mixed-aerobic athletes (mean age, 24.2 ± 6.5 years, body mass index, 22.2 ± 1.2) were studied after giving informed consent. These athletes included five international class cross-country skiers, four national class middle- and long-distance runners, one semicompetitive distance runner, one rower, and one national class basketball player. The study was performed with the approval of the Human Ethics Committee of the University of Tartu (Tartu, Estonia).

Protocol for exercise

Subjects underwent a maximal exercise test to determine maximal O_2 consumption (VO_{2max}). They performed an incremental treadmill (LE 3000) exercise test until volitional exhaustion. VO_{2max} was measured using "breath by breath" gas monitoring (Oxycon Record, Erich Jaeger, Germany). One week later, all subjects exercised for 60 min at 80% of their VO_{2max} after a 5-min warm-up. Subjects were asked to refrain from intensive exercise for at least 3 days preceding the test. On the day of exercise testing, the subjects ate a light carbohydrate-rich breakfast. The exercise tests were carried out 2–4 h after breakfast.

Blood sampling

Blood was drawn from an antecubital vein before and immediately after sustained exercise at 80% VO_{2max} .

Routine blood analyses

Complete blood cell counts, including total leukocyte, neutrophil, lymphocyte, and monocyte counts, hemoglobin, and hematocrit were measured from EDTA-treated blood using an automated hematology analyzer (Sysmex SE9000, Japan).

Determination of lymphocyte subpopulations

Flow cytometry was performed to determine the lymphocyte subpopulations (12). Whole blood leukocytes were labeled with monoclo-

nal antibodies against surface determinants to identify the $CD3^+$ (T-cell), $CD19^+$ (B-cell), and $CD16^+CD56^+/CD3^-$ (natural killer cell) lymphocyte subpopulations. All antibodies were purchased from Becton–Dickinson (San Jose, CA, U.S.A.). Data were acquired and analyzed on a FACSort flow cytometer by two-color flow cytometric analysis using SimulSET software (Becton–Dickinson, Palo Alto, CA, U.S.A.).

Tumor necrosis factor- α (TNF α) and interleukin 2 receptor (IL-2R) assays

IMMULITE TNF α and IMMULITE IL-2R solid-phase, two-site chemiluminescent enzyme immunometric assays (DPC, U.K.) for use with the IMMULITE Automated Analyzer were used for the quantitative measurement of plasma TNF α and soluble plasma IL-2R (1, 4, 5).

Lymphocyte isolation and assay of NF- κ B activation

Lymphocytes were isolated from fresh whole blood using Ficoll-Paque PLUS (Pharmacia, 17-1440-02) and low-speed centrifugation as described by others (8). The isolated suspension of lymphocytes was subjected to whole-cell protein extraction for the determination of NF- κ B as described before (27). In brief, total cell extracts were prepared using a high-salt detergent buffer. Lymphocytes were resuspended and lysed in 0.050 ml of buffer containing 20 mM HEPES, pH 7.5, 400 mM NaCl, 1 mM $MgCl_2$, 0.5 mM EDTA, 20% glycerol, 1% Nonidet P-40, 0.5 mM dithiothreitol, 0.010 mg/ml leupeptin, 1 mM phenylmethylsulfonyl fluoride, and 0.001% aprotinin. The extracts were centrifuged for 15 min at 14,000 rpm in a microcentrifuge at 4°C, and clear supernatant was stored at -70°C. Protein concentration in the supernatant was determined using the Bradford protein assay reagent (Bio-Rad, Hercules, CA, U.S.A.). The samples were subjected to electrophoretic mobility shift assay for the determination of NF- κ B activity (27).

Binding reaction mixtures contained 0.02 mg of extracted protein, 0.002 mg of ds+sDNA as nonspecific competitor, 0.002 mg of bovine serum albumin, 20 mM HEPES, pH 7.5, 50 mM NaCl, 0.5 mM $MgCl_2$, 2% glycerol, and 0.25 ng of ^{32}P -end-labeled NF- κ B specific double-

stranded oligonucleotide probe (Santa Cruz Biotechnology Inc., Santa Cruz, CA, U.S.A.). To determine the specificity of the NF- κ B band, a competition assay (13) was performed using a 20-fold excess of the unlabeled probe. To assess the subunit composition of DNA binding protein, specific antibodies were used for supershift assay (13). Cellular extracts were incubated with antibodies against p65 (Santa Cruz Biotechnology Inc.) subunits of NF- κ B for 15 min at room temperature before the binding reaction mixture was added. After the binding reaction (30 min at room temperature), the samples were run on a native 4.8% polyacrylamide gel in 0.5 \times TBE buffer (12.5 mM Tris-borate containing 0.25 mM Na₂EDTA, pH 8.0). NF- κ B bands were detected by autoradiography and quantified by densitometry (13).

Lipid peroxidation markers

Serum conjugated diene (CD) levels were measured according to methods previously described (25) with minor modifications (35). In brief, 0.25% butylated hydroxytoluene (BHT)-treated serum samples (0.15 ml) and 0.15 ml of 0.9% NaCl were incubated at 37°C for 25 min, and lipids were extracted by heptane/isopropanol (1:1). Samples were acidified by 5 mol/L hydrochloric acid and extracted by cold heptane. After centrifugation for 5 min at 3,000 rpm, the absorbance of the heptane fraction was measured at the absorbance maximum between 220 and 250 nm using isotonic saline as blanks.

Conversion of conjugated dienes results in stable secondary products (aldehydes, alkenals) of lipid peroxidation. The assay for thiobarbituric acid (TBA) reactive products (TBARS), in which TBA reacts with malondialdehyde, is a sensitive but relatively nonspecific method for detection of later stage lipid peroxidation. TBARS levels were detected in serum samples (23) with minor modifications to increase specificity (35). In brief, samples were treated with BHT twice, immediately after collection and before addition of the test reagents, to suppress oxidative changes during handling and analysis. Hemolyzed samples were excluded from analysis. Samples (0.25 ml) were incubated with 0.475 mM Fe²⁺ at 37°C for

30 min. After incubation, BHT (0.25%) was added to the samples. This mixture was treated with acetate buffer, pH 3.5, and heated with TBA solution (1%, 80°C, 70 min). The samples were then cooled and acidified (5 mol/L hydrochloric acid). After extraction with cold butanol, samples were centrifuged. The absorbance of the butanol fraction was measured at 534 nm. A standard plot for malondialdehyde (end-product of lipid peroxidation) was prepared using 1,1,3,3-tetraethoxypropane.

Statistical analysis

All results are presented as means \pm SD. TNF α and TBARS levels were log-transformed before statistical analyses to correct for skewing, although raw values are presented in the results. ANOVA for repeated measures was used for analyzing the change of variables with exercise. Results were adjusted as necessary for hemoconcentration by using changes in hematocrit with exercise as a covariate. In all analyses, a value of $p < 0.05$ was considered statistically significant.

RESULTS

The mean VO_{2max} (68.6 \pm 7.6 ml/min/kg) indicated that the men were highly fit. Both the hemoglobin concentration and hematocrit rose slightly with exercise (Table 1, $p = 0.035$ – 0.039).

Sixty minutes of strenuous cycling exercise induced moderate leukocytosis in the peripheral blood stream (Table 1). Neutrophilia was most prominent ($p = 0.003$). The total peripheral lymphocyte count tended to increase ($p = 0.058$), but not after controlling for plasma volume contraction as estimated by the change in hematocrit. On the other hand, the natural killer cell lymphocyte subpopulation showed a marked increase with exercise ($p = 0.007$). Monocyte levels remained unchanged.

Physical exercise caused activation of NF- κ B in eight of 12 individuals. No appreciable change was detected in three men, and in one case (subject 4, Fig. 1) preexercise baseline activity was higher than postexercise activity (Fig. 1). Supershift assay using anti-p65 antibody (lanes 1 and 2 from left; Fig. 1A) showed

TABLE 1. PERIPHERAL WHITE BLOOD CELL COUNTS, PLASMA LEVELS OF TNF- α , SOLUBLE IL2-R, AND LIPID PEROXIDATION IN 12 YOUNG MEN JUST BEFORE AND JUST AFTER 60 MIN OF EXERCISE AT 80% VO_{2MAX}

	Preexercise	Postexercise	p
Hemoglobin (g/L)	153 \pm 13.04	160 \pm 9.0	0.035
Hematocrit (%)	0.435 \pm 0.04	0.45 \pm 0.02	0.039
Leukocytes			
Total white blood cells (cells \times 10 ⁹ /L)	5.56 \pm 1.11	8.10 \pm 2.15	0.001
Lymphocytes (cells \times 10 ⁹ /L)	1.49 \pm 0.48	1.76 \pm 0.38	0.058
T-cells (cells \times 10 ⁹ /L)	1.05 \pm 0.36	1.14 \pm 0.29	0.404
B-cells (cells \times 10 ⁹ /L)	0.174 \pm 0.05	0.192 \pm 0.02	0.440
Natural killer cells (cells \times 10 ⁹ /L)	0.26 \pm 0.16	0.44 \pm 0.23	0.007
Neutrophils (cells \times 10 ⁹ /L)	3.23 \pm 0.81	5.60 \pm 2.11	0.003
Monocytes (cells \times 10 ⁹ /L)	0.57 \pm 0.16	0.55 \pm 0.15	0.670
Cytokine and receptor			
TNF α (pg/ml)	21.8 \pm 6.3	25.7 \pm 5.2	0.003
Soluble IL-2R (U/ml)	543 \pm 174	577 \pm 195	0.005
Lipid peroxidation			
TBA reactivity (μ M malondialdehyde equivalents)	1.63 \pm 0.38	1.93 \pm 0.28	0.009
Conjugated dienes (μ M)	50.94 \pm 10.14	56.39 \pm 10.86	0.047

Data are means \pm SD. See Materials and Methods for statistical treatment.

that the NF- κ B protein complex contained the transcriptionally active p65 protein. When the one case of exceptional outlying response (subject 4, Fig. 1B) was excluded from analyses, the mean NF- κ B activity increased by >50% ($p = 0.002$) as detected from densitometry data.

Strenuous exercise increased plasma TNF α ($p = 0.003$) and soluble plasma IL-2R ($p = 0.005$) levels (Table 1). These increases were significant even after controlling for changes in hematocrit. Plasma conjugated diene ($p = 0.047$) and especially TBARS ($p = 0.009$) concentrations increased with exercise (Table 1).

DISCUSSION

In humans, physical exercise markedly influences immune function (34, 39). In physically fit men, strenuous exercise for 1 h increased total white blood cell count in the peripheral blood. In 1893, Schultz first described that exercise may cause leukocytosis (26). Several studies have repeated this observation (39). Physical exercise is also known to influence the composition of the lymphocyte subset in peripheral blood (9, 10, 18, 19). The nature of this effect is thought to be dependent

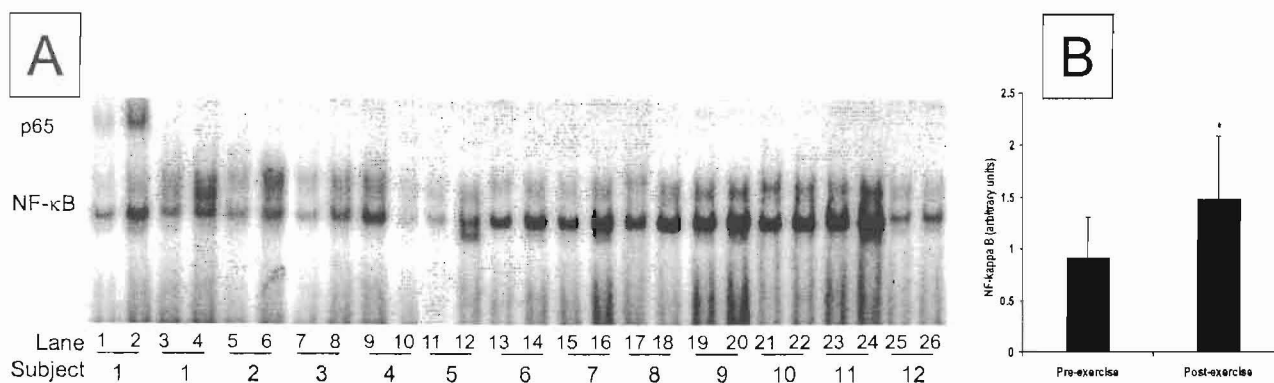


FIG. 1. DNA-binding of NF- κ B harvested from peripheral blood lymphocytes of 12 young men before and after 60 min of exercise at 80% VO_{2max} . Lanes 1 and 2, supershift evidence; lanes 3 and 4, preexercise and postexercise from subject 1, respectively; lanes 5 and 6, subject 2; etc. The first two lanes show supershift data for the presence of p65. (B) Densitometry data. Units are arbitrary, with preexercise levels set at 1.0. p for the difference between before and after exercise was 0.061. After omission of an outlying response (subject 4), the difference was highly significant ($p = 0.002$).

on the exercise type and intensity. Exercise-induced changes in the lymphocyte subset pattern are not related to changes in blood volume, suggesting that a bout of exercise directly influences lymphocyte function and fate (15). In response to the exercise test used in the current study, no significant changes were observed in total lymphocyte or monocyte count, T-cell count, or B-cell count. Consistent with previous reports (20), the count of natural killer lymphocytes in the peripheral blood markedly increased in response to exercise. An increase in plasma epinephrine during exercise is thought to be one factor responsible for the exercise-induced increase in peripheral blood natural killer cell count (24). Neutrophilia is known to occur in response to endurance exercise (36). We observed that the 1-h bout of exercise markedly enhanced the peripheral blood neutrophil count. Exercise-induced changes in peripheral blood cell counts observed in this study represented typical changes that are expected in response to endurance exercise.

Results of this study provide first evidence that strenuous exercise in well trained young men may cause activation of NF- κ B. The NF- κ B activation process is known to be transient (2, 3, 7, 30). Activated NF- κ B signals for the expression of the inhibitor protein I κ B. I κ B, thus expressed, terminates NF- κ B activation and resets the NF- κ B activation switch in the cytosol (2, 3, 7, 30). This transient nature of NF- κ B activation may explain the lack of exercise-induced NF- κ B activity in three of 12 men. Perhaps the NF- κ B response kinetics in these men were such that they were not detectable at the time point the samples were collected. Basal NF- κ B activity is usually low, and the activity of this transcription factor is known to be induced by a wide variety of stimuli (2, 3, 7, 30). Consistently, we have observed that NF- κ B activity in preexercise baseline samples was low in 11 of 12 subjects. In the one exceptional case, a factor unknown to us contributed to high baseline NF- κ B activity. Consistent with the electrophoretic mobility shift assay, physical exercise also enhanced the expression of NF- κ B-directed gene products TNF α and IL2-R (2, 3, 7, 30). This effect was observed in each and every subject studied (individual data not shown). This line of evidence is consistent with

our observation that physical exercise-induced activation of NF- κ B proteins includes the transcriptionally active p65 protein. It is therefore evident that exercise-induced cytosolic activation of NF- κ B is associated with a transactivation response. The results presented, however, do not allow delineation of the mechanism of exercise-induced NF- κ B activation. It is possible, however, that the effect is triggered by a combination of multiple factors. In this study it was observed that exercise caused oxidative stress in the blood as evident from lipid peroxidation data. At the same time, exercise elevated plasma levels of TNF α . Both reactive oxygen species and TNF α are thought to be potent activators of NF- κ B (14, 21, 30, 33). It has been also demonstrated that TNF α signaling may involve reactive oxygen species as intracellular messengers (17, 37, 38). It is thus of interest to examine whether exercise-induced NF- κ B activation is inhibited by dietary antioxidant supplementation.

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ABBREVIATIONS

BHT, butylated hydroxytoluene; CD, conjugated diene; IL-2R, interleukin-2 receptor; NF- κ B, nuclear factor- κ B; TBA, thiobarbituric acid; TBARS, TBA reactive substances; TNF α , tumor necrosis factor- α ; VO $_{2\max}$, maximal O $_2$ consumption.

REFERENCES

1. Babson AL, Olson DR, Palmieri T, Ross AF, Becker DM, and Mulqueen PJ. The IMMULITE assay tube: a new approach to heterogeneous ligand assay. *Clin Chem* 37: 1521-1522, 1991.
2. Baeuerle PA and Henkel T. Function and activation

- of NF-kappa B in the immune system. *Annu Rev Immunol* 12: 141–179, 1994.
3. Baldwin AS Jr. The NF-kappa B and I kappa B proteins: new discoveries and insights. *Annu Rev Immunol* 14: 649–683, 1996.
 4. Costongs GM, van Oers RJ, Leerkes B, and Janson PC. Evaluation of the DPC IMMULITE random access immunoassay analyser. *Eur J Clin Chem Clin Biochem* 33: 887–892, 1995.
 5. Einarsson R, Lei JD, Ullrich A, and Van Dalen A. Performance characteristics of IMMULITE TPS: a comparison with TPS IRMA [In Process Citation]. *Anticancer Res* 19: 2743–2747, 1999.
 6. Gerondakis S, Grumont R, Rourke I, and Grossmann M. The regulation and roles of Rel/NF-kappa B transcription factors during lymphocyte activation. *Curr Opin Immunol* 10: 353–359, 1998.
 7. Ghosh S, May MJ, and Kopp EB. NF-kappa B and Rel proteins: evolutionarily conserved mediators of immune responses. *Annu Rev Immunol* 16: 225–260, 1998.
 8. Harris R and Ukaejiofo EO. Tissue typing using a routine one-step lymphocyte separation procedure. *Br J Haematol* 18: 229–235, 1970.
 9. Hoffman-Goetz L. Influence of physical activity and exercise on innate immunity. *Nutr Rev* 56: S126–S130, 1998.
 10. Hoffman-Goetz L, Simpson JR, Cipp N, Arumugam Y, and Houston ME. Lymphocyte subset responses to repeated submaximal exercise in men. *J Appl Physiol* 68: 1069–1074, 1990.
 11. Inoue T, Mu Z, Sumikawa K, Adachi K, and Okochi T. Effect of physical exercise on the content of 8-hydroxydeoxyguanosine in nuclear DNA prepared from human lymphocytes. *Jpn J Cancer Res* 84: 720–725, 1993.
 12. Jackson A. Basic phenotyping of lymphocytes: selection and testing reagents and interpretation of data. *Clin Immunol Newslett* 10: 43–55, 1990.
 13. Janssen YM and Sen CK. Nuclear factor kappa B activity in response to oxidants and antioxidants. *Methods Enzymol* 300: 363–374, 1999.
 14. Kaltschmidt B, Sparna T, and Kaltschmidt C. Activation of NF- κ B by reactive oxygen intermediates in the nervous system. *Antioxid Redox Signal* 2: 129–144, 1999.
 15. Kargotich S, Keast D, Goodman C, Crawford GP, and Morton AR. The influence of blood volume changes on leucocyte and lymphocyte subpopulations in elite swimmers following interval training of varying intensities. *Int J Sports Med* 18: 373–380, 1997.
 16. Karin M. The NF-kappa B activation pathway: its regulation and role in inflammation and cell survival. *Cancer J Sci Am* 4(Suppl 1): S92–S99, 1998.
 17. Li Y-P, Atkins CP, Sweatt JD, and Reid MB. Mitochondria mediate tumor necrosis factor alpha/ NF- κ B signaling in skeletal muscle myotubes. *Antioxid Redox Signal* 1: 97–104, 1999.
 18. Mars M, Govender S, Weston A, Naicker V, and Chuturgoon A. High intensity exercise: a cause of lymphocyte apoptosis? *Biochem Biophys Res Commun* 249: 366–370, 1998.
 19. Mazzeo RS, Rajkumar C, Rolland J, Blaher B, Jennings G, and Esler M. Immune response to a single bout of exercise in young and elderly subjects. *Mech Ageing Dev* 100: 121–132, 1998.
 20. Moyna NM, Acker GR, Weber KM, Fulton JR, Robertson RJ, Goss FL, and Rabin BS. Exercise-induced alterations in natural killer cell number and function. *Eur J Appl Physiol* 74: 227–233, 1996.
 21. Muller JM, Rupec RA, and Baeuerle PA. Study of gene regulation by NF-kappa B and AP-1 in response to reactive oxygen intermediates. *Methods* 11: 301–312, 1997.
 22. Niess AM, Baumann M, Roecker K, Horstmann T, Mayer F, and Dickhuth HH. Effects of intensive endurance exercise on DNA damage in leucocytes. *J Sports Med Phys Fitness* 38: 111–115, 1998.
 23. Ohkawa H, Ohishi N, and Yagi K. Reaction of linoleic acid hydroperoxide with thiobarbituric acid. *J Lipid Res* 19: 1053–1057, 1978.
 24. Palmo J, Asp S, Daugaard JR, Richter EA, Klokke M, and Pedersen BK. Effect of eccentric exercise on natural killer cell activity. *J Appl Physiol* 78: 1442–1446, 1995.
 25. Recknagel RO and Glende EA Jr. Spectrophotometric detection of lipid conjugated dienes. *Methods Enzymol* 105: 331–337, 1984.
 26. Schultz G. Experimentelle Untersuchungen über das Vorkommen und die diagnostische Bedeutung der Leucocytose. *Dtsch Arch Klin Med* 51: 234–281, 1893.
 27. Schulze-Osthoff K, Beyaert R, Vandevoorde V, Haegeman G, and Fiers W. Depletion of the mitochondrial electron transport abrogates the cytotoxic and gene-inductive effects of TNF. *EMBO J* 12: 3095–3104, 1993.
 28. Schulze-Osthoff K, Los M, and Baeuerle PA. Redox signalling by transcription factors NF-kappa B and AP-1 in lymphocytes. *Biochem Pharmacol* 50: 735–741, 1995.
 29. Sen CK. Oxidants and antioxidants in exercise. *J Appl Physiol* 79: 675–686, 1995.
 30. Sen CK and Packer L. Antioxidant and redox regulation of gene transcription [see comments]. *FASEB J* 10: 709–720, 1996.
 31. Sen CK, Packer L, and Hanninen O. *Exercise and Oxygen Toxicity*. Amsterdam: Elsevier Science Publishers B.V, 1994, p. 536.
 32. Sen CK, Packer L, and Hanninen O. *Handbook of Oxidants and Antioxidants in Exercise*. Amsterdam: Elsevier, 2000, p. 1189.
 33. Sen CK, Sies H, and Baeuerle PA. *Antioxidant and Redox Regulation of Genes*. San Diego: Academic Press, 2000, p. 556.
 34. Shephard RJ and Shek PN. Immune responses to inflammation and trauma: a physical training model. *Can J Physiol Pharmacol* 76: 469–472, 1998.
 35. Starkopf J, Zilmer K, Vihalemm T, Kullisaar T, Zilmer M, and Samarutel J. Time course of oxidative stress

- during open-heart surgery. *Scand J Thorac Cardiovasc Surg* 29: 181–186, 1995.
36. Suzuki K, Naganuma S, Totsuka M, Suzuki KJ, Mochizuki M, Shiraishi M, Nakaji S, and Sugawara K. Effects of exhaustive endurance exercise and its one-week daily repetition on neutrophil count and functional status in untrained men. *Int J Sports Med* 17: 205–212, 1996.
37. Wong GH and Goeddel DV. Induction of manganous superoxide dismutase by tumor necrosis factor: possible protective mechanism. *Science* 242: 941–944, 1988.
38. Wong GH, Elwell JH, Oberley LW, and Goeddel DV. Manganous superoxide dismutase is essential for cellular resistance to cytotoxicity of tumor necrosis factor. *Cell* 58: 923–931, 1989.
39. Woods JA, Davis JM, Smith JA, and Nieman DC. Exercise and cellular innate immune function. *Med Sci Sports Exerc* 31: 57–66, 1999.

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